



Isolation of Lactic Acid Bacteria as a Potential Probiotic in Dangke, a Traditional Food from Enrekang, Indonesia

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Abstract

This research is a preliminary study to explore potential probiotics of Lactic Acid Bacteria (LAB) in dangke, a traditional food from cow milk in Enrekang, Indonesia. Isolation of LAB performed using the selective medium de Man Rogosa Sharpe Agar. LAB will show clear zone on MRS medium after the addition of the indicator in the form of CaCO₃ 1% and incubated for 24 hours. Selection is done by observing the cell morphology and Gram staining. Further testing with the biochemical properties of sugar fermentation test. To assess their probiotic ability, tolerance of low pH were examined. The results showed that isolates of lactic acid bacteria detected were genera of Lactobacillus and could survive at pH 2. Isolates of Lactobacillus in dangke are derived from cow milk showed potential as a candidate probiotic bacteria.

Keywords: Lactic Acid Bacteri; Lactobacillus; Dangke, Probiotic.

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1. Introduction

Probiotic bacteria are defined FAO/WHO as 'Live microorganisms which when administered in adequate amounts confer a health benefit on the host' [1]. The research of invitro and invivo, toward experimental animal or human, has been conducted many times to explore the role of probiotic in prevention or therapy of disease, for instance diarrhea [2], Inflammatory Bowel Disease [3], Irritable Bowel Syndrome [4, 5], asthma [6, 7], allergy [8, 9], obesity [10, 11], type 2 of diabetes [12, 13], colorectal cancer [14-16], intolerant lactose [17, 18], depression [19-21], osteoporosis [22, 23], reduction of cholesterol level [24, 25], and binding of heavy metal ion [26, 27].

The role of probiotic to prevention and therapy of some diseases has been proved, but its price is still relatively expensive. Therefore, the searching of new probiotic source needs to do. Research about isolation of probiotic potential bacteria has been conducted in many times such as from adult humans feces [28], infant feces [29], pickles of Kyoto [30], kefir [25], kimchi [31], and sake [32]. Some other researches conducted bacteria isolation from milk and milk processing, such as from ASI [33], cow milk [34], buffalo milk [35], horse milk [36], fermented milk [37], dadih [38], and dangke [39].

Dangke is milk processing product of buffalo or cow traditionally which comes from South Sulawesi especially the district of Enrekang. Dangke is processed by being heated with small fire until it boils, then it is added coagulant such as papaya sap (papain) so there will be clotting. The clot is taken into special mold made of coconut shell while being pushed so the liquid gets separated [40].

The aim of this research is to explore potential probiotics of Lactic Acid Bacteria (LAB) in dangke, a traditional food from cow milk in Enrekang, Indonesia. In this research, LAB isolation is conducted by using selective medium de Man Rogosa Sharpe Agar, observing the cell morphology, Gram staining, biochemical properties test, and testing of bacteria viability on low pH.

2. Material and Methods

Sample Preparation of Dangke

There was 10 gram dangke suspended into sterile physiological liquid (NaCl 0.9%) as many as 90 ml and it was taken into homogene. From the suspension, it was taken 1 ml and put into thinner tube which contained 9 ml sterile aquades, it was taken into homogene producing dilution of 10^{-1} . The dilution was continued until dilution of 10^{-3} .

Isolation and Selection of Lactic Acid Bacteria from Dangke

Dangke suspension inoculated to liquid medium of MRS Broth then it was incubated for 24 hours by temperature 37°C . The culture of MRS Broth was inoculated to MRSA medium then it was added CaCO_3 1%, next it was incubated for 48 hours. The colony around it was formed transparent zone purified again on MRSA medium by continuous scratch method then it was incubated for 24 – 48 hours. Kultivation was conducted many

times over on the same medium and condition until we got single colony. Then the pure isolate was moved to slant agar as stock, it was kept in refrigerator by temperature 4°C.

Bacteria Identification by Gram Staining and Biochemical Test

The pure isolate was taken aseptically and put on object glass cleaned by alcohol 96% and it was given fixation on spiritus lamp. After cold, it was dropped Gram A (Violet Crystal) 2-3 drops for 1 minute, then it was cleaned by flowing water and dried in the air. By the same procedure, the was continued by Gram B (Iodium) for 1 minute, Gram C (Alcohol 96 %) for 30 seconds, and Gram D (Safranin) for 45 seconds. This observation was conducted by looking at the form and color of cell under microscope with certain magnification.

Biochemical test that was conducted comprised KIA test, Motility, Catalase by putting the pure isolate on the object glass then it was dropped H₂O₂, it was observed if there was gas bubble produced or not, and Carbohydrate test (glucose, lactose, sucrose, maltose, manitol and malonat) by taking in the pure isolate one ose in every medium and it was taken into homogene then incubated by temperature 37°C for 1 x 24 hour.

Tolerance of Low pH

One ose of pure isolate was inoculated into 5 ml medium of MRS Broth, incubation on temperature 37°C for 24 hours. Then 1% inoculum was taken into 5 ml MRS Broth which was managed its pH by using HCl with variation pH 2,0; 2,5; 3,0; 3,5 and 4,0. Incubation was conducted with temperature 37°C for 24 hours. Counting the number of bacteria in inoculum at the beginning and the last of incubation was conducted by plating method using MRSA.

3. Results and Discussion

Isolation and Selection of Lactic Acid Bacteria from Dangke

There were three isolates derived from dangke cow milk which grew on medium of MRSA + CaCO₃ 1% for 1 x 24 hours by temperature 37° C which had different colony morphology comprised form, side, elevation, and color. The different colony morphology became basic for assumption that the three isolates were different kinds of bacteria. After being conducted the purification for the three isolates and being conducted the screening by using medium of MRSA added with 1% CaCO₃ as a medium used to select LAB, two isolates showed transparent zone around its colony as a consequence of being produced lactic acid reacted with CaCO₃ forming Ca-lactic which fused in medium [33].

Each of two selected isolates was isolate A and B. Isolate A had small colony, medium in round form, convex elevation, flat side, bright surface, and milk white color. While isolate B showed different colony morphology with isolate A, namely medium colony to big and round colony, flat side, bright surface, and milk white color. The different from isolate A was on the bigger colony size.

Bacteria Identification by Gram Staining and Biochemical Test

Next two selected isolates were observed its Gram characteristics and biochemical activities to concern about characterization and identification.

Table 1: Biochemical test of Lactic Acid bacteria

Biochemical Test	Bacteria Isolates	
	A	B
Gram Staining	Bacil of Positive Gram	Bacil of Positive Gram
Catalase	-	-
KIA	A/A, -/-	A/A, -/-
Urea	-	-
S. Citrat	-	-
LIA	-	-
MIO	-/ /-	-/ /-
Glucose	+	+
Lactose	+	-
Sucrose	+	+
Maltose	+	+
Manitol	+	-
Malonat	-	-
	Lactobacillus A	Lactobacillus B

Gram characteristics and biochemical activities on table 1 which is suited for Bergey’s Manual of Determinative Bacteriology shows that isolate are group of LAB identified as Lactobacillus [41]. Lactobacillus is rod-shaped, often long and slender. Non-motile. Gram-positive. Pigment production rare; when present, yellow or orange to rust or brick-red. Gelatin is not liquefied. Growth on potato is poor or absent. Glucose and similar aldehydic hexoses, carbohydrates which yield these simple sugars, and polyhydroxy alcohols are changed either by homofermentation to lactic acid or by heterofermentation to lactic and acetic acids, alcohol and carbon dioxide. Nitrates are not reduced except under certain conditions with Lactobacillus plantarum. Several species grow at relatively high temperatures. Poor surface growth because these bacteria are generally microaerophilic or anaerobic. Do not produce catalase. Found in fermenting animal (especially dairy) and plant products [41].

Tolerance of Low pH

In this research, Lactobacillus A and Lactobacillus B were tested their growing ability on low pH, namely pH 2.0, 2.5, 3.0, 3.5, and 4.0. Test of bacteria resistance to low pH is one of the most important characteristics in determining bacteria potential to be probiotic [1]. The result of resistance of LAB isolate toward low pH can be seen on Table 2 and 3.

Results showed that based on House Index (HI), Container index (CI) and Breteau index (BI), the most

successful program to decrease larva index of Aedes aegypti mosquitoes was abate program following by fogging and health education.

Table 2: The Number of Bacteria Cell of Lactobacillus A on Low pH

Variation of pH	The First Number of Bacteria Cell (CFU/ml)	The Last Number of Bacteria Cell (CFU/ml)
2.0	1.3×10^8	2.5×10^6
2.5	1.3×10^8	5.6×10^6
3.0	1.3×10^8	8.6×10^7
3.5	1.3×10^8	7.4×10^8
4.0	1.3×10^8	8.6×10^9

Table 3: The Number of Bacteria Cell of Lactobacillus B on Low pH

Variation of pH	The First Number of Bacteria Cell (CFU/ml)	The Last Number of Bacteria Cell (CFU/ml)
2.0	1.1×10^8	8.5×10^6
2.5	1.1×10^8	1.4×10^7
3.0	1.1×10^8	6.6×10^8
3.5	1.1×10^8	9.3×10^8
4.0	1.1×10^8	7.6×10^8

Lactobacillus isolated from dangke of cow milk shows that the ability to survive on low pH. Both of isolates can grow with mortality level less than 3 log so it can be stated that the isolate is resistant on acid state. Each bacteria cell of Lactobacillus A and Lactobacillus B grew on pH 2 with amount 2.5×10^6 CFU/ml (there was a decreasing of cell number of 2 log) and 8.5×10^6 CFU/ml (there was a decreasing of cell number of 2 log). The number of first inoculum each Lactobacillus A and Lactobacillus B added was 1.3×10^8 CFU/ml and 1.1×10^8 CFU/ml. The beginning LAB used for probiotic test was 10^8 to 10^9 CFU/ml. In acid state, Lactobacillus can maintain its cytoplasmic acidity so that the protein and enzyme in cell can keep working optimally. Isolate of LAB can adapt on low pH because it has regulation system of cell internal pH (pHi). This case can be achieved with new enzymes synthesis and it produces proton (H+) from inside of cell of which release happens through

hydrolysis process of ATP (H⁺-ATPase). LAB hold out from damage of acid because there are histidin dekarboksilase and arginin deiminasi enzyme. The tolerance of LAB toward acid is high enough because its ability to maintain sitoplasm pH has more bases than extraseluler pH [42]. Lactobacillus A and Lactobacillus B are positive Gram bacteria which have resistance toward the very extreme pH [40]. Both of bacteria can pass an interior cavity of stomach producing acid so that it can achieve down intestines to be able to push pathogen bacteria which likely stay in digestive line. Bacteria will have effect on intestine environment if the number of population of the bacteria achieves minimal 10⁶ – 10⁸ CFU/ml. Based on the result of tolerant test of bacteria isolate from dangke cow milk toward low pH, it showed that both of isolates have potential as candidate probiotic bacteria

4. Conclusion

This research showed that Lactic Acid Bacteria isolated from dangke, a traditional food from cow milk in Enrekang, Indonesia is Gram positive, rod shaped, non motile. The results showed that isolates of lactic acid bacteria detected were genera of Lactobacillus and could survive at pH 2. Isolates of Lactobacillus in dangke are derived from cow milk showed potential as a candidate probiotic bacteria.

References

- [1]. Araya M, Morelli L, Reid G, Sanders ME, Stanton C. 2002. Report of a Joint FAO/WHO Working Group on Drafting Guidelines for the Evaluation of Probiotics in Food, London Ontario, Canada.
- [2]. Guandalini and Stefano MD. 2011. Probiotics for Prevention and Treatment of Diarrhea. *Journal of Clinical Gastroenterology*. 45:149-153
- [3]. Rauch M and Lynch S. 2012. The potential for probiotic manipulation of the gastrointestinal microbiome. *Curr Opin Biotechnol*. 23(2):192-201
- [4]. Ringel Y and Ringel-Kulka T. 2011. The rationale and clinical effectiveness of probiotics in irritable bowel syndrome. *J Clin Gastroenterol*. 45:145-8
- [5]. Ki CB, Mun JS, Hwan CC, Song ID, Woong LH, Joon KH, Hyuk J, Kyung CS, Kim K, Chung WS, Seo JG. 2012. The effect of a multispecies probiotic mixture on the symptoms and fecal microbiota in diarrhea-dominant irritable bowel syndrome: a randomized, double-blind, placebo-controlled trial. *J Clin Gastroenterol*. 46(3):220-7
- [6]. Jan RL, Yeh KC, Hsieh MH, Lin YL, Kao HF, Li PH, Chang YS, Wang JY. 2011. Lactobacillus gasseri suppresses Th17 pro-inflammatory response and attenuates allergen-induced airway inflammation in a mouse model of allergic asthma. *Br J Nutr*. 14:1-10
- [7]. Jang SO, Kim HJ, Kim YJ, Kang MJ, Kwon JW, Seo JH, Kim HY, Kim BJ, Yu J, Hong SJ. 2012. Asthma Prevention by Lactobacillus Rhamnosus in a Mouse Model is Associated With CD4+CD25+Foxp3+ T Cells. *Allergy Asthma Immunol Res*. 4(3):150-156
- [8]. Drago L, Iemoli E, Rodighiero V, Nicola L, De Vecchi E, Piconi S. 2011. Effects of Lactobacillus salivarius LS01 (DSM 22775) treatment on adult atopic dermatitis: a randomized placebo-controlled study. *Int J Immunopathol Pharmacol*. 24(4):1037-48
- [9]. Wang J, Zhang H, Chen X, Chen Y, Menghebilige, Bao Q. 2012. Selection of potential probiotic

- lactobacilli for cholesterol-lowering properties and their effect on cholesterol metabolism in rats fed a high-lipid diet. *J Dairy Sci.* 95(4):1645-54
- [10]. Luoto R, Kalliomäki M, Laitinen K, Isolauri E. 2010. The impact of perinatal probiotic intervention on the development of overweight and obesity: follow-up study from birth to 10 years. *Int J Obes.* 34(10):1531-1537
- [11]. Arora T, Anastasovska J, Gibson G, Tuohy K, Sharma RK, Bell J, Frost G. 2012. Effect of *Lactobacillus acidophilus* NCDC 13 supplementation on the progression of obesity in diet-induced obese mice. *Br J Nutr.* 31:18
- [12]. Ejtahed HS, Mohtadi NJ, Homayouni RA, Niafar M, Asghari-Jafarabadi M, Mofid V. 2012. Probiotic yogurt improves antioxidant status in type 2 diabetic patients. *Nutrition.* 28(5):539-43
- [13]. Andersson U, Bränning C, Ahrné S, Molin G, Alenfall J, Onning G, Nyman M, Holm C. 2010. Probiotics lower plasma glucose in the high-fat fed C57BL/6J mouse. *Benef Microbes.* 1(2):189-96
- [14]. Kumar RS, Kanmani P, Yuvaraj N, Paari KA, Pattukumar V, Thirunavukkarasu C, Arul V. 2012. *Lactobacillus plantarum* AS1 isolate from south Indian fermented food Kallappam suppress 1,2-dimethyl hydrazine (DMH)-induced colorectal cancer in male Wistar rats. *Appl Biochem Biotechnol.* 166(3):620-31
- [15]. Chen CC, Lin WC, Kong MS, Shi HN, Walker WA, Lin CY, Huang CT, Lin YC, Jung SM, Lin TY. 2011. Oral inoculation of probiotics *Lactobacillus acidophilus* NCFM suppresses tumour growth both in segmental orthotopic colon cancer and extra-intestinal tissue. *Br J Nutr.* 30:1-12
- [16]. Appleyard CB, Cruz ML, Isidro AA, Arthur JC, Jobin C, De Simone C. 2011. Pretreatment with the probiotic VSL#3 delays transition from inflammation to dysplasia in a rat model of colitis-associated cancer. *Am J Physiol Gastrointest Liver Physiol.* 301(6):1004-13
- [17]. Ojetti V, Gigante G, Gabrielli M, Ainora ME, Mannocci A, Lauritano EC, Gasbarrini G, Gasbarrini A. 2010. The effect of oral supplementation with *Lactobacillus reuteri* or tilactase in lactose intolerant patients: randomized trial. *Eur Rev Med Pharmacol Sci.* 14(3):163-70
- [18]. He T, Priebe MG, Zhong Y, Huang C, Harmsen HJ, Raangs GC, Antoine JM, Welling GW, Vonk RJ. 2008. Effects of yogurt and bifidobacteria supplementation on the colonic microbiota in lactose-intolerant subjects. *J Appl Microbiol.* 104(2):595-604
- [19]. Arseneault BJ, Rondeau I, Gilbert K, Girard SA, Tompkins TA, Godbout R, Rousseau G. 2011. Combination of *Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175 reduces post-myocardial infarction depression symptoms and restores intestinal permeability in a rat model. *Br J Nutr.* 21:1-7
- [20]. Desbonnet L, Garrett L, Clarke G, Kiely B, Cryan JF, Dinan TG. Effects of the probiotic 2010. *Bifidobacterium infantis* in the maternal separation model of depression. *Neuroscience.* 170(4):1179-88
- [21]. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, Bienenstock J, Cryan JF. 2011. Ingestion of *Lactobacillus* strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc Natl Acad Sci U S A.* 108(38): 16050–16055
- [22]. Yeo SK and Liong MT. 2010. Angiotensin I-converting enzyme inhibitory activity and bioconversion of isoflavones by probiotics in soymilk supplemented with prebiotics. *Int J Food Sci Nutr.* 61(2):161-81

- [23]. Rodrigues FC, Castro AS, Rodrigues VC, Fernandes SA, Fontes EA, de Oliveira TT, Martino HS, Ferreira CL. 2012. Yacon Flour and *Bifidobacterium longum* Modulate Bone Health in Rats. *J Med Food*.
- [24]. Kumar M, Nagpal R, Hemalatha R, Verma V, Kumar A, Chakraborty C, Singh B, Marotta F, Jain S, Yadav H. 2012. Cholesterol-lowering probiotics as potential biotherapeutics for metabolic diseases. *Exp Diabetes Res*. 2012:902-917
- [25]. Wang HF, Tseng CY, Chang MH, Lin JA, Tsai FJ, Tsai CH, Lu YC, Lai CH, Huang CY, Tsai CC. 2012. Anti-inflammatory effects of probiotic *Lactobacillus paracasi* on ventricles of BALB/C mice treated with ovalbumin. *Chin J Physiol*. 55(1):37-46
- [26]. Buda B, Dylus E, Górska-Frączek S, Brzozowska E, Gamian A. 2013. Biological properties of *Lactobacillus* surface proteins. *Postepy Hig Med Dosw*. 67: 229-237
- [27]. Zhai Q, Wang G, Zhao J, Liu X, Tian F, Zhang H, and Chen W. 2013. Protective Effects of *Lactobacillus plantarum* CCFM8610 against Acute Cadmium Toxicity in Mice. *Appl Environ Microbiol*. 79(5): 1508–1515.
- [28]. Tinrat S, Saraya Sumam, and Chomnawang MT. 2011. Isolation and Characterization of *Lactobacillus salivarius* MTC 1026 as a Potential Probiotic. *J. Gen. Appl. Microbiol*. 57:365-378
- [29]. Chiang SS and Pan TM. 2012. Beneficial effects of *Lactobacillus paracasei* subsp. *paracasei* NTU 101 and its fermented products. *Applied Microbiology and Biotechnology*. 93(3):903-16
- [30]. Nonaka Y, Izumo T, Izumi F, Maekawa T, Shibata H, Nakano A, Kishi A, Akatani K, Kiso Y. 2008. Antiallergic effects of *Lactobacillus pentosus* strain S-PT84 mediated by modulation of Th1/Th2 immunobalance and induction of IL-10 production. *Int Arch Allergy Immunol*. 145(3):249-57
- [31]. Won TJ, Kim B, Lim YT, Song DS, Park SY, Park ES, Lee DI, Hwang KW. 2011. Oral administration of *Lactobacillus* strains from Kimchi inhibits atopic dermatitis in NC/Nga mice. *J Appl Microbiol*. 110(5): 1195-202
- [32]. Kawamoto S, Kaneoke M, Ohkouchi K, Amano Y, Takaoka Y, Kume K, Aki T, Yamashita S, Watanabe K, Kadowaki M, Hirata D, Ono K. 2011. Sake Lees Fermented with Lactic Acid Bacteria Prevents Allergic Rhinitis-Like Symptoms and IgE-Mediated Basophil Degranulation. *Biosci. Biotechnol. Biochem*. 75(1), 140-144
- [33]. Djide MN dan Wahyudin E. 2008. Isolasi Bakteri Asam Laktat dari Air Susu Ibu, dan Potensinya dalam Penurunan Kadar Kolesterol Secara In Vitro. *Majalah farmasi dan Farmakologi*. 12(3):73-78
- [34]. Baruzzi F, Poltronieri P, Quero GM, Morea M, Morelli L. 2011. An in vitro protocol for direct isolation of potential probiotic lactobacilli from raw bovine milk and traditional fermented milks. *Appl Microbiol Biotechnol*. 90(1):331-42
- [35]. Duary RK, Rajput YS, Batish VK, Grover S. 2011. Assessing the adhesion of putative indigenous probiotic lactobacilli to human colonic epithelial cells. *Indian J Med Res*. 134(5): 664–671
- [36]. Shi T, Nishiyama K, Nakamata K, Aryantini NPD, Mikumo D, Oda Y, Yamamoto Y, Mukai T, Sujaya IN, Urashima T, and Fukuda K. 2012. Isolation of Potential Probiotic *Lactobacillus rhamnosus* Strains from Traditional Fermented Mare Milk Produced in Sumbawa Island of Indonesia. *Biosci. Biotechnol. Biochem*. 76(10):1879-1903
- [37]. Vizoso PMG, Schuster T, Briviba K, Watzl B, Holzapfel WH, Franz CM. 2007. Adhesive and

- chemokine stimulatory properties of potentially probiotic *Lactobacillus* strains. *Journal of Food Protection*. 70(1):125-34
- [38]. Surono IS, Koestomo FP, Novitasari N, Zakaria FR, Yulianasari, Koesnandar. 2011. Novel probiotic *Enterococcus faecium* IS-27526 supplementation increased total salivary sIgA level and bodyweight of pre-school children: a pilot study. *Anaerobe*. 17(6):496-500
- [39]. Razak AR, Patong AR, Harlim T, Djide MN, Haslia, Mahdalia. 2009. Produksi Senyawa Bakteriosin Secara Fermentasi Menggunakan Isolat BAL *Enterococcus faecium* DU55 dari Dangke. *Indonesia Chemica Acta*. 2(2):19
- [40]. Marzoeki, A.A.M., A. Hafid, M., Jufri, Amir, Madjid. 1978. Penelitian Peningkatan Mutu Dangke. Balai Penelitian Kimia Departemen Perindustrian, Makassar.
- [41]. Breed RS, Murray EGD, Smith NR. 1957. *Bergey's Manual of Determinative Bacteriology*. 7th ed. Baltimore. The Williams & Wilkins Company.
- [42]. Pan DD, Zeng XQ, Yan YT. 2011. Characterisation of *Lactobacillus fermentum* SM-7 isolated from koumiss, a potential probiotic bacterium with cholesterol-lowering effects. *J Sci Food Agric*. 91(3):5128.
- [43]. Holt JG. 1994. *Bergey's Manual of Determinative Bacteriology*. 9th ed. ISBN-10: 0683006037, 13: 978-0683006032. Lippincott. Williams & Wilkins Company.