



Determination of the Effect of the Environment on the Genetic Polymorphism In the Genus of *Tamarix* Using the Molecular Marker (Simple Sequence Repeats "PCR-SSR" (In Arid Areas of the Khenchela Region (Eastern of Algeria)

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Abstract

Tamarix is used for a long time as a treatment plant in traditional and modern medicine because of its healing powers. In addition to these medicinal characteristics *Tamarix* has a great adaptation to the extreme environmental conditions, especially in arid and semi arid regions, where it represents a promising source for rehabilitation of degraded ecosystems and fight against desertification. However, to date, only limited information about its genetic structure. To attempt to characterize the genetic diversity of the species of this genus is therefore very useful for their classification, their conservation and improvement.

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In this context, we started in the assessment of the level of genetic diversity characterizing *Tamarix* pushing currently in Algeria and in which they are originate. we used the PCR-SSR of 23 genotypes selected from three very different sites (en perspective of climate, edaphic and orographic conditions), through the arid regions of Khenchela in eastern of Algeria. The results are obtained by the highlighted amplification of 14 primers specific for *Tamarix*. Analysis of the results using the software TreeCon for Windows, at a threshold of dissimilarity of 50% we have obtained nine distinct groups, in general, the results showed a high level of genetic divergence within and between species among genotypes, with relevant remarks in four accessions, (II 2, II 10, III 3 and III 9), of the ninth group who have not shown any genetic variation linked to the species or to the environment.

Keywords: *Tamarix*; arid regions; genetic polymorphism; PCR-SSR; Khenchela; Algeria

1. Introduction

The *Tamarix* is one of four genera of the family of *Tamaricaceae*, containing more than 80 species whose are native to Eurasia and North of Africa [1]. There is more than 14 species and subspecies of *Tamarix* in Algeria whom are determined in the new flora of Algeria Quezel [2]. The *Tamarix* has been known since antiquity for its therapeutic characteristics, and is even used as an anti-carcinogen plant in traditional medicine among the peoples of the Maghreb [3].

The *Tamarix* is distinguished by its high ubiquity and ecological resistance, where it can tolerate to the most of extreme environmental conditions such as drought, salinity, cold etc.. [4]. Those characteristics make from *Tamarix* a promising shrubby plant for the rehabilitation of ecosystems and the fight against desertification in arid and semi-arid, where Algeria including more than 80% of the lands [5].

However, the mechanisms of adaptation and genetic structure of the genus of *Tamarix* are not known, which is the case of many Mediterranean species, few studies have brought on the assessment of genetic diversity, which is the necessary knowledge for good management and development of the plant genetic resources [6]. This study contributes to the identification of the effect of the environment on the genetic variability of the genus of *Tamarix*. To do so we opted for the technique molecular marker (PCR-SSR) of 23 accessions of different species of the genus of *Tamarix*, using 14 specific primers for *Tamarix*. The genotypes were collected from three very different sites in climatic, edaphic and orographic levels and distributed along a south-north transect in arid zone of Khenchela which is located in eastern Algeria.

2. Materials and Methods

2.1. Plant material and test sites

Samples of *Tamarix* species studied were collected from three different sites (Site I (Shott), Site II (Oued (River)) Site III (Desert)) from the arid zone of Khenchela in eastern Algeria (Figure 1 and 2). We selected at random 30 samples consisting of young leaves of 10 adult individuals for each site.

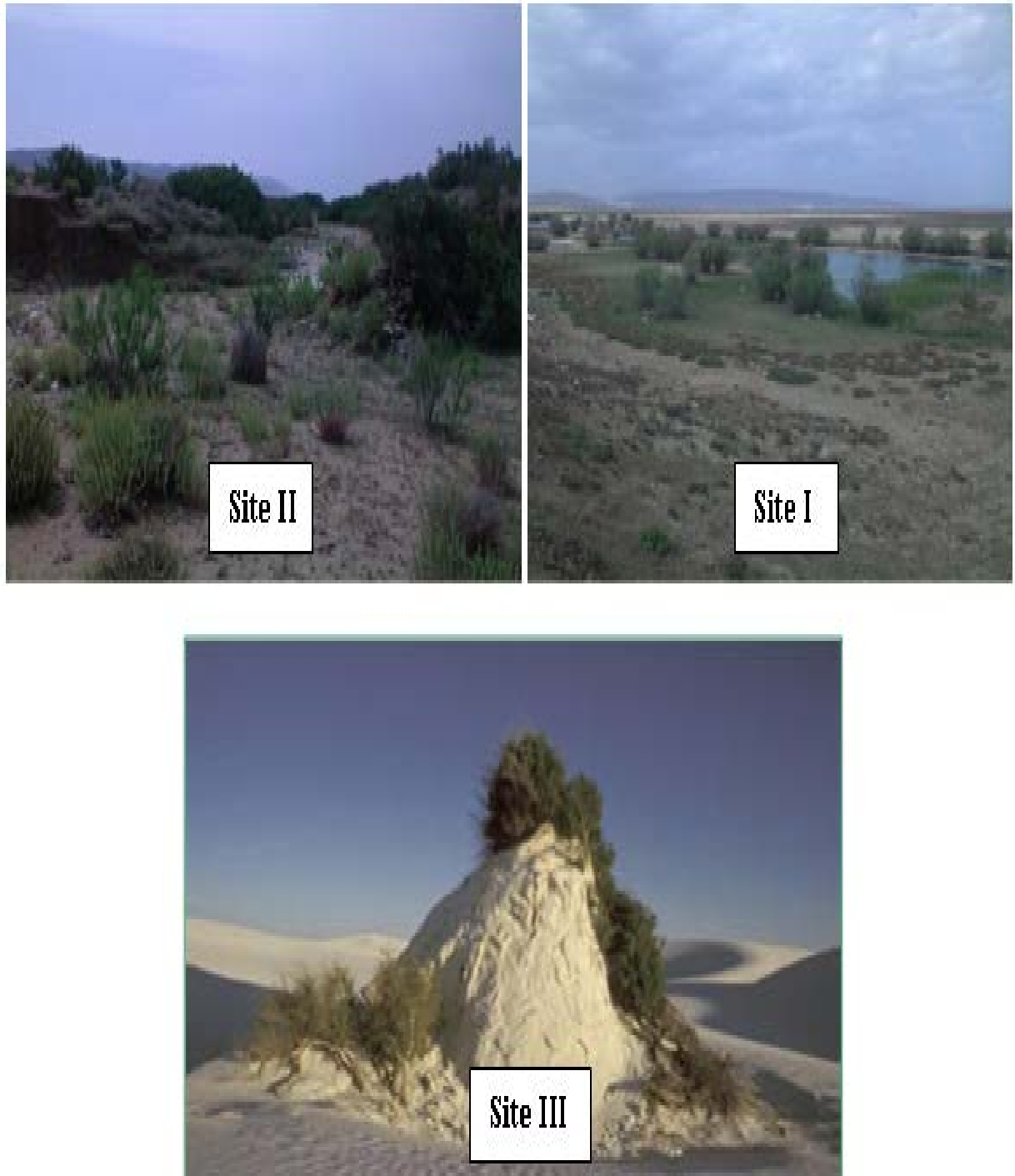


Figure 1: Panoramic view of the study's sites

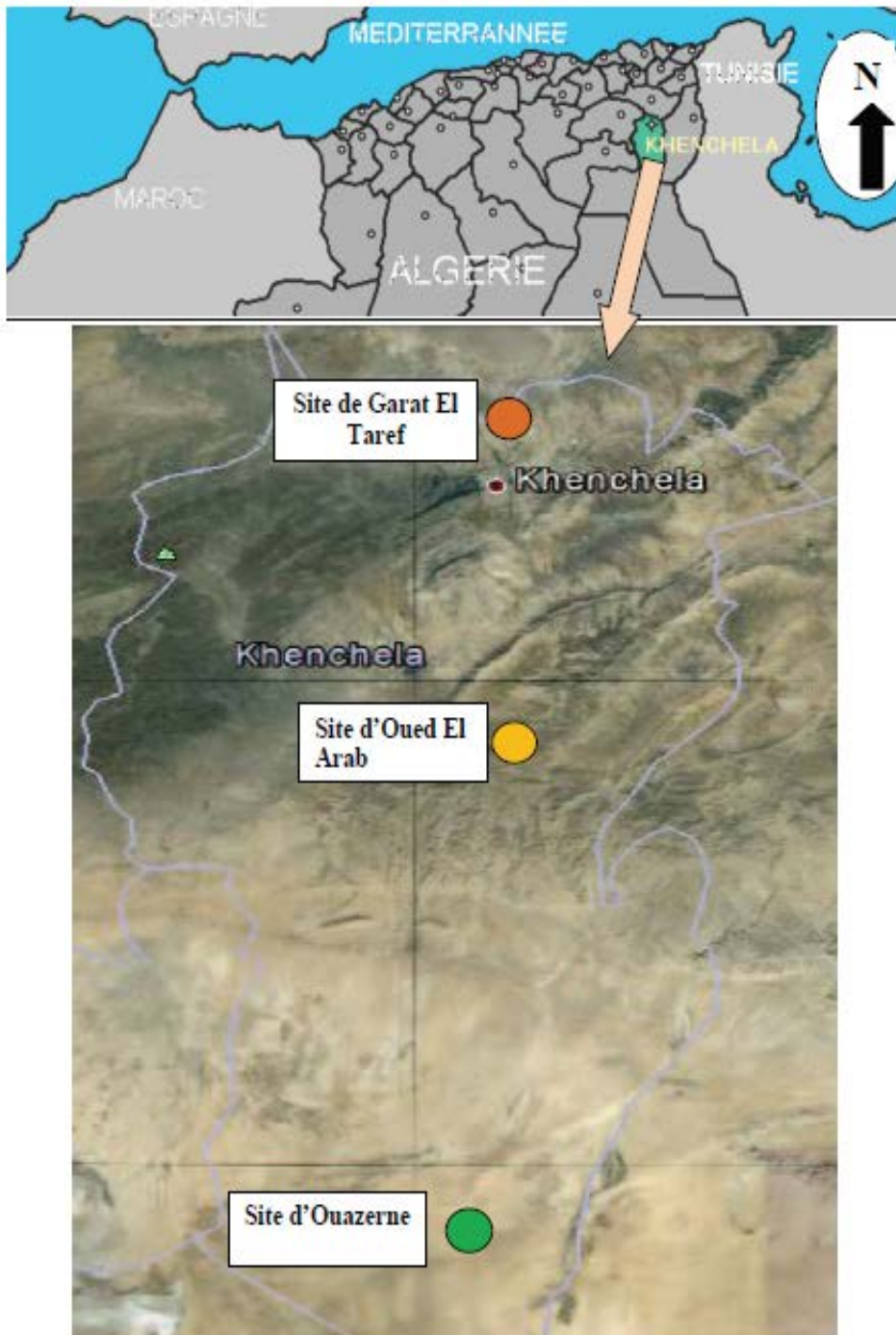


Figure 2: The geographical position of the region of Khenchela and distribution of study's sites

2.2. Extraction and quantification of the DNA

Genomic DNA was extracted from young leaves using the cetyltrimethylammonium bromide (CTAB), by grinding in liquid nitrogen, using the method described by [7,8]. The different samples purified DNA was resuspended in 100 μ l of TE buffer solution. The DNA concentration was monitored by quantification performed on 0.8% agarose gel.

2.3. Primers used

We tested 30 genotypes with 14 specific primers of *Tamarix*, because of the quality of polymorphic bands they produced, we selected 23 genotypes whom gave a good results (Table 01).

Table 1: the primers used and genotypes selected to SSR markers

Primer	Séquence	Repeat Motif	Ta (C °)	Good results' Génotypes		
				Site I	Site II	Site III
Th127	F:TTGGCTGTTGAAGAAGATCG	(AGG) 4	58	I 1,	II 1,	III 3,
	R:TCTCCAAACCTTGACCGACT			I 2,	II 2,	III 4,
Th321	F:TACCTTGCGAACACAACACTGC	(CTC) 6	58	I 3,	II 3,	III 5,
	R:TACACCGAGAGAGACGCTGA					
Ta201	F:AATTTGTCCGACTCCACTGT R:CGTCTCCTTTTCAGGCGTAG	(AT) 7	56	I 4,	II 4,	III 6,
Th412	F:CTGGCAAGTAGCAACACCTCT R:GGATGAACAACCCAACCATC	(AG) 11	58	I 5,	II 6,	III 8,
Th715	F:ACGTGGTTTGGTGAAAGGAG	(AG) 10	54	I 6,	II 8,	III 9,
	R:CCACCCTTAACCCACTCAGA			I 7,	II 10.	III 10.
Th1071	F:CGCTCTGTTGATCATCTTCG	(TTTC) 4	58	I 8,		
	R:TGTCCCAATCCGTTACAAAA					
Ta1350	F:CATGGCAGTGATGGATTGAG R:GGACAGTTCAGCCTCCACAT	(GA) 7	57	I 9.		
Th2620	F:GTTGAGCAGCAATCACATGC R:GAAGGGGCAGTGTTTTTCAA	(AAG) 6	58			
Th2876	F:CTGTAGCCAAGCATGGGACT R:AAGACACGTAAACCCGCAAC	(CCTGCT)4	58			
Th3484	F:TCAGATTTTGCAAACCACCA R:AAGCCTTTGCATACCACCAC	(CATC) 4	58			
Th5990	F:GCCGAATTTTGTGTTGGATT R:AATAAAAGGCACCCTCATCG	(ATG) 11	57			
Th6387	F:TCGGATTCTGGAAGGTGTTT R:TGCAACGAAAACATTATTACCC	(TTA) 6	54			
Th6976	F:CCGTGGACTAACCTTGCCTA R:CAAGCAAACGCAGGGTAGAT	(ATG) 11	58			

2.4. DNA amplification and electrophoresis

Amplification was performed in a total volume of 25µl according to the Protocol of Gaskin [9], [10] containing:
- 2µl of DNA extract (10ng/µl);

- Buffer 5µl enzyme polymerization (5x);
- 2µl of MgCl₂ (2.5 mM);
- 2.5µl dNTPs (2 mM);
- 3.8µl oligonucleotide primer (2µM);
- 0.4µl of Taq DNA polymerase (5U/µl).
- the remainder of the volume is completed with bidistilled water up to 25µl.

The PCR process consists of an initial pre-denaturation (3 minutes at 95 ° C), followed by 45 cycles of amplification (30 sec denaturation at 95 ° C, 1 min annealing at 37 ° C, 2 min extension at 72 ° C) and a final extension of 10 min at 72 ° C [11].

The PCR products were separated by agarose gel electrophoresis in 2% (w / v TBE) containing ethidium bromide for 90 min. The DNA bands are then observed under ultraviolet light and photographed using GelDoc (Figure 3) .

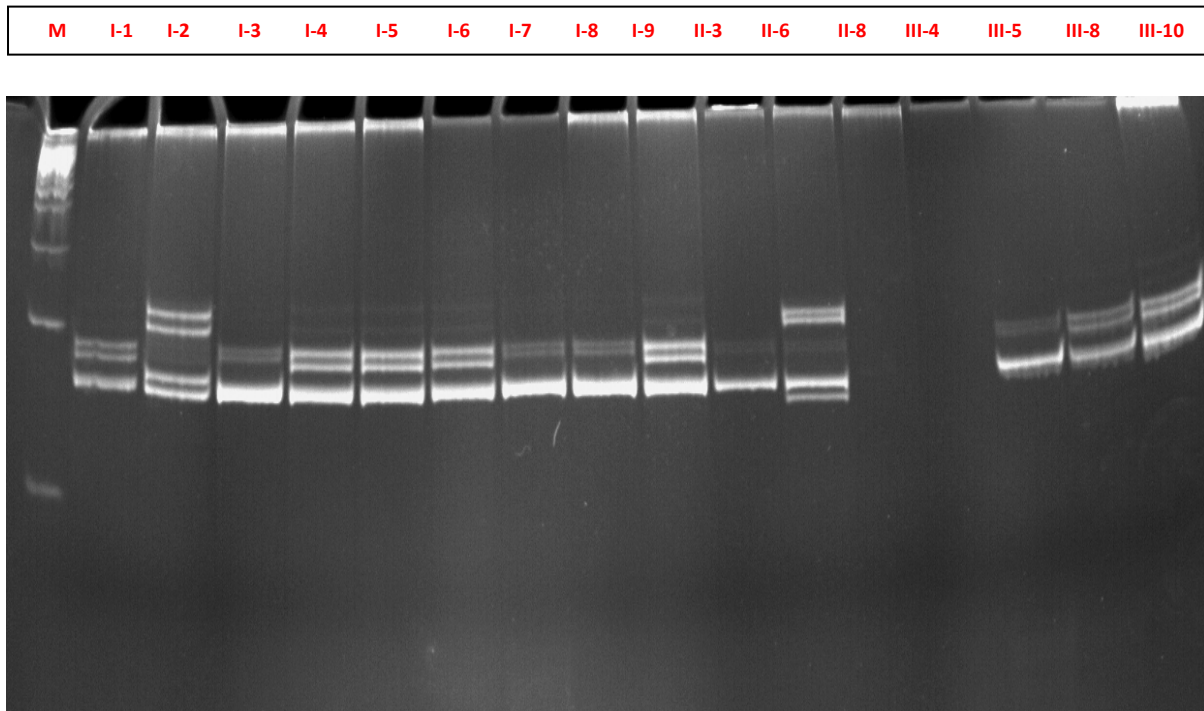


Figure 3: Example of the result obtained with the primer Th412:Ta = 58 ° C, Taq = 0.4µl, DNA =2µl

2. 5. Bioinformatic analysis of data

For each individual and for each primer, the visual reading of the gels allowed to note polymorphic bands, we assigned the value 1 for the band present and 0 for absent band. After receiving the corresponding data in the binary matrix obtained in text format file extension (*. Txt) in Notepad, the TreeCon For windows software has developed a tree using the NJ method (Neighbor-joining) and obtaining a matrix of coefficients of genetic similarity between different genotypes studied of *Tamarix* (Table 2).

Table 2: Matrix of similarity between the genotypes of *Tamarix* using the Neighbor Joining method (legend: in red; remotest accessions, in black; remote accessions and in blue; nearest accessions)

I-1	0																						
I-2	58,62	0																					
I-3	52,73	49,3	0																				
I-4	50,94	50,73	45,46	0																			
I-5	36,17	58,73	50	48,27	0																		
I-6	53,19	61,91	60	51,72	26,92	0																	
I-7	46,34	61,4	62,96	76,92	47,83	60,87	0																
I-8	54,39	53,43	42,86	61,77	58,07	64,52	46,43	0															
I-9	42,37	46,67	47,22	42,86	46,88	53,13	51,72	43,24	0														
II-1	91,3	89,74	94,44	88,24	85,71	85,71	90,91	89,47	95	0													
II-2	90,91	94,74	94,29	93,94	92,59	92,59	90,48	94,6	94,87	33,33	0												
II-3	60	73,91	72,93	85,37	71,43	82,86	51,72	68,89	65,96	81,82	80	0											
II-4	92	85,37	78,95	88,89	86,67	86,67	75	90	85,71	66,67	60	84,62	0										
II-6	52,38	34,18	39,47	48,65	50	52,94	48,39	41,03	40	95,46	95,35	64,71	82,61	0									
II-8	75	90	83,78	88,57	79,31	86,21	91,3	94,87	95,12	60	50	83,33	71,43	91,11	0								
II-10	90,91	94,74	94,29	93,94	92,59	92,59	90,48	94,6	94,87	33,33	80	60	95,35	50	0	0							
III-3	90,91	94,74	94,29	93,94	92,59	92,59	90,48	94,6	94,87	33,33	80	60	95,35	50	0	0	0						
III-4	62,16	69,81	52	79,17	52,38	71,43	61,11	53,85	59,26	88,89	88,24	68	80	68,97	78,95	88,24	88,24	0					
III-5	72,97	69,81	68	70,83	66,67	67,57	1466,67	53,85	66,67	77,78	88,24	68	90	72,41	89,47	88,24	88,24	56,25	0				
III-6	82,61	94,87	88,89	88,24	85,71	85,71	81,82	89,47	90	50	33,33	63,64	66,67	90,91	60	33,33	33,33	88,89	77,78	0			
III-8	68,42	66,67	60,78	75,51	72,09	62,79	72,97	62,26	60	89,47	88,89	61,54	80,59	72,88	90	88,89	88,89	39,39	51,52	89,47	0		
III-9	90,91	94,74	94,29	93,94	92,59	92,59	90,48	94,6	94,87	33,33	80	60	95,35	50	0	0	88,24	88,24	33,33	88,89	0		
III-10	53,57	50	47,83	64,18	60,66	57,38	63,64	49,3	50,69	94,6	94,44	68,18	89,74	45,46	84,21	94,44	94,44	56,86	56,86	89,19	38,46	94,44	0
I-1	I-2	I-3	I-4	I-5	I-6	I-7	I-8	I-9	II-1	II-2	II-3	II-4	II-6	II-8	II-10	III-3	III-4	III-5	III-6	III-8	III-9	III-10	

3. Results and Discussion

The results obtained allowed us to distribute the 23 genotypes into nine distinct groups, at 50% of similarity, according to the dendrogram produced by the TreeCon for Windows software (Figure 4):

- The first group (G1) comprises accessions: I-1, I-9, I-4, I-5 and I-6, all belonging to the site I, and they can be divided into 3 subgroups with a threshold 45 % similarity.

- the first subgroup includes accessions I-1 and I-9 with a similarity coefficient of 42.37. The second subgroup includes the accession I-4 alone .The third subgroup includes I-5 and I-6 with a coefficient of 26.92. All these accessions are genetically close, the division of this group into three subgroups can be explained by the effect of the environment in which some genotypes are emerged in salt water (Shott) and other genotypes are so far and they can be a subspecies of *Tamarix gallica* L.
- The second group (G2) includes accessions I-2, I-3, I-8 and II-6 it can also be divided into 2 subgroups, first subgroup I-2 and II-6 with a coefficient of 34.18 of similarity, and the second subgroup includes the accessions I-3 and I-8 with a coefficient of 42.86 of similarity. This genetic combination may be explained by the fact that these accessions have the same adaptation, same height, same branching

twigs and even flowering time that we let that thought to the genotypes of *Tamarix chottica* Trab. which characterizes these environments (Quezel, 1963).

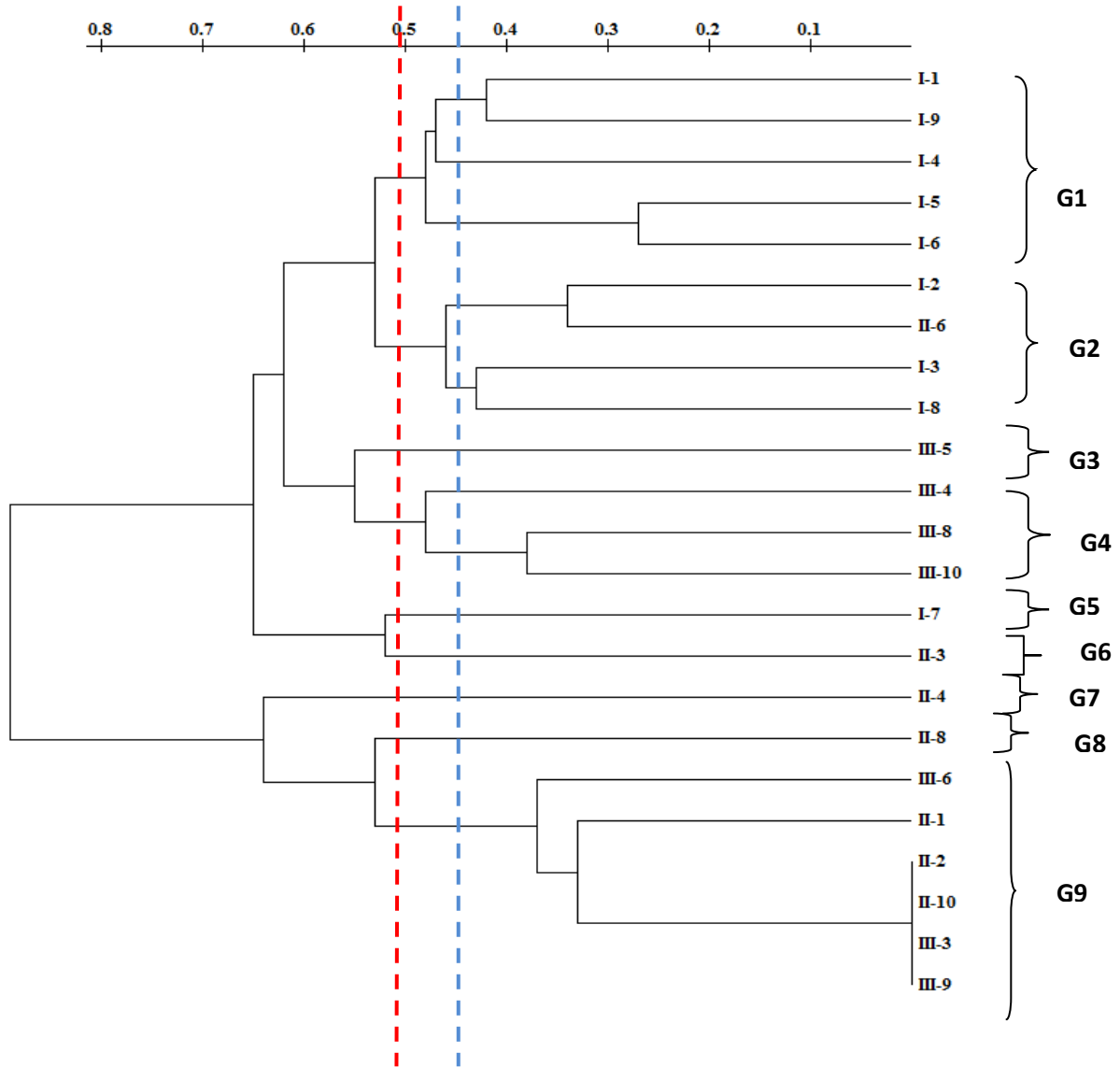


Figure 4: Dendrogram representing genetic distances existing between the genotypes studied generated by SSR markers. (Based on the dissimilarity coefficient by using the Neighbor Joining method (NJ)). The groups (G1 to G9) are shown at the right of FIG. The vertical red dotted line indicates a groups' separation at 50% of similarity and The vertical blue dotted line indicates the groups' at 45% of dissimilarity.

- The third group (G3) contains only the accession III-5 indicates that it wants a quite different genotype.
- The fourth group (G4) includes accessions III-4, III-8 and III-10 where the accession III-4 constitutes a first subgroup ,while accessions III-8 and III-10 constitute a second subgroup this approximation can

be allows us to judge that these accessions belong to the *Tamarix parviflora* in Ehrenb.

- The fifth group (G5): includes only the accession I-7 .
- The sixth group (G6): includes only the accession II-3.
- The seventh group (G7): includes only the accession II-4.
- The 8th group (G8) includes only the accession II-8.

All those groups which are represented by a single accession for each, this means that each accession represents a distinct species from other.

The 9th group (G9) includes accessions :III-6, II-1, II-8, II-10, III-3 and III-9 including the last 4 are closely related although both sites II and III are more than 120 km away from each other, probably, it is the same accession *Tamarix africana* Poir. which is multiplied vegetatively in both sites.

4. Conclusion

The results allow us to confirm the phenotypic polymorphism in the *Tamarix* which is a major problem to botanists for the classification of species and subspecies of this kind, where we noticed that the accessions studied are genetically very distant because in most cases the dissimilarity coefficients are greater than 50%, which explains a large genetic polymorphism between and within specific in *Tamarix*.

However, we found that accessions are closer despite the distance between them, as in the case of group 9, which eliminates the effect of the environment on polymorphism in this case. It should be noted that the interpretation of some cases of separation and / or genetic merger are hotly debated. By way of perspective, and the advances in genetic engineering, *Tamarix* may have, no doubt, a better characterization of their genetic structure. That is why it is imperative to use more sophisticated methods of genetic investigation, ranging from the use of more specific markers to the identification, sequencing and gene transfer of interest. Thus, and involving the workforce more representative genotypes, it will happen, eventually, to get the maximum genetic information, leading to better management and greater value of this phyto-genetic resources that are shrubby plants form the genus of *Tamarix* which will become important in the restoration of arid and hyper arid regions.

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